

KRAS, BRAF, BRCA1 and EGFR Mutation-Specific Panels using Hydrogel-Based 3D *In Vitro* Tumor Models

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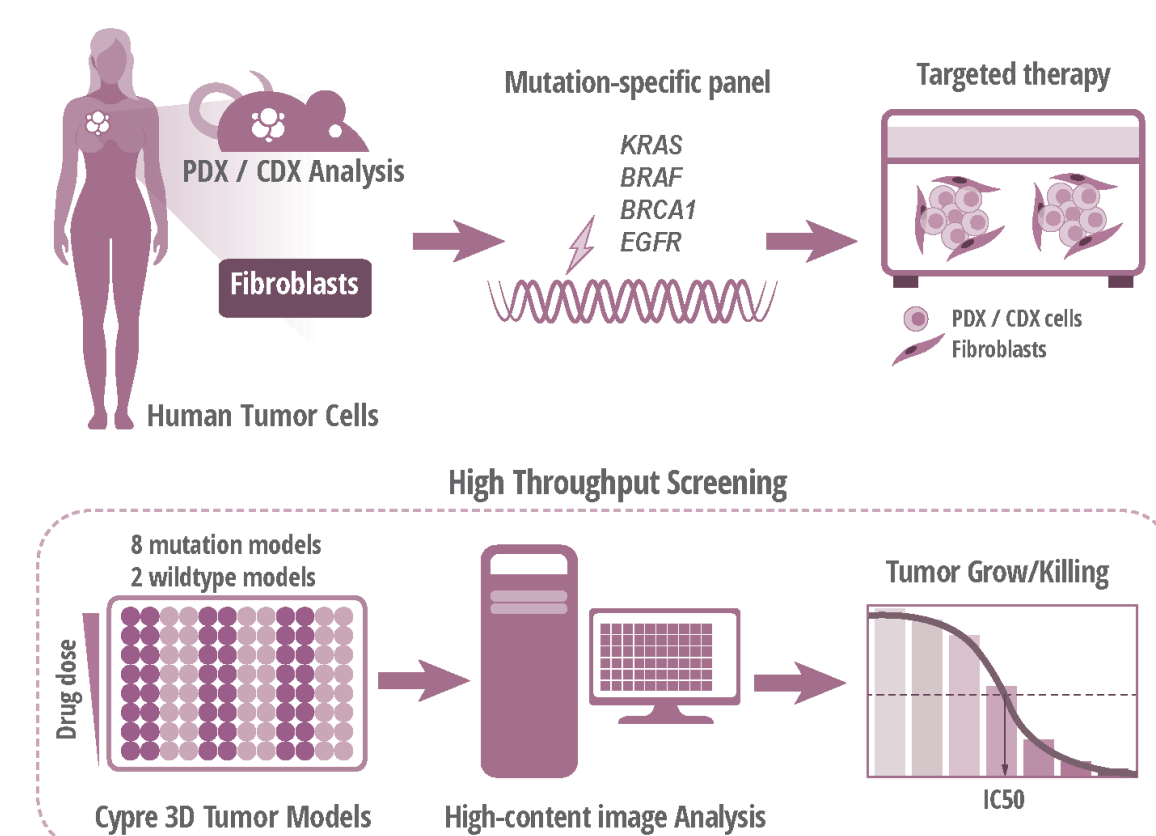
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1 ABSTRACT

Mutation-targeting therapies have become a transformative technology and, in many ways, a new paradigm for cancer treatment for patients. The early development of Erlotinib for non-small cell lung cancer paved the way for EGFR treatment, irrespective of tumor indication subtype. And the more recent launch of Sotorasib for *KRAS*^{G12C} mutation has marked a turning point for the once undruggable mutation. The advent of gene specific-treatments requires a reconfiguration of preclinical models which more effectively represent the said mutation in populations of diverse backgrounds and tumor indications. 3D models which have the ability to recreate the key hallmarks of cancer - such as growth, invasion, immune infiltration and suppression, stromal transformation, drug diffusion, as well as gene mutational signatures - are of exceptional interest to drive rapid and scalable compound efficacy screening.

Here, we developed four 3D *in vitro* tumor panels carrying mutations in one of the following genes - KRAS, EGFR, BRCA1, and BRAF - as well as wildtype models for comparison. The tumors were grown in a hydrogel matrix in 96-well plates with stromal fibroblasts and screened with the standard of care (e.g. Sotorasib for *KRAS*^{G12C}, Erlotinib for *EGFR*^{mut}, the PARP inhibitor Olaparib for *BRCA*^{mut}, and Vemurafenib for *BRAF*^{V600E}) and other known therapeutic compounds that target their respective mutations. The utility of a 3D hydrogel matrix and fibroblasts in the panels' models were key in recreating the stromal compartment of the tumor microenvironment. Endpoints were determined within four days of treatment, including tumor size reduction and tumor killing, via high-content image analysis. The Panels' robustness as demonstrated by the reproducible results in independent experiments suggests they can be utilized at *in vitro* pharmacology stages and earlier in Hit-to-lead screens.

2 PLATFORM



A streamlined workflow for growing 3D patient-derived xenograft tumors (PDX) or cell-line derived xenograft (CDX) models and assaying targeted drugs using high content analysis and advanced analytics. The standardized setup includes 96-well format and 6-dose in duplicates, and assay endpoints include tumor size and cell death using high content imaging.

**Visit: <https://www.criver.com/cancer-model-database> to learn the PDX lines in the panel.

1 RESULTS

Fig 1. Dose-response analysis of the 3D tumor growth/killing of Sotorasib and Vemurafenib in Cypre 3D platform

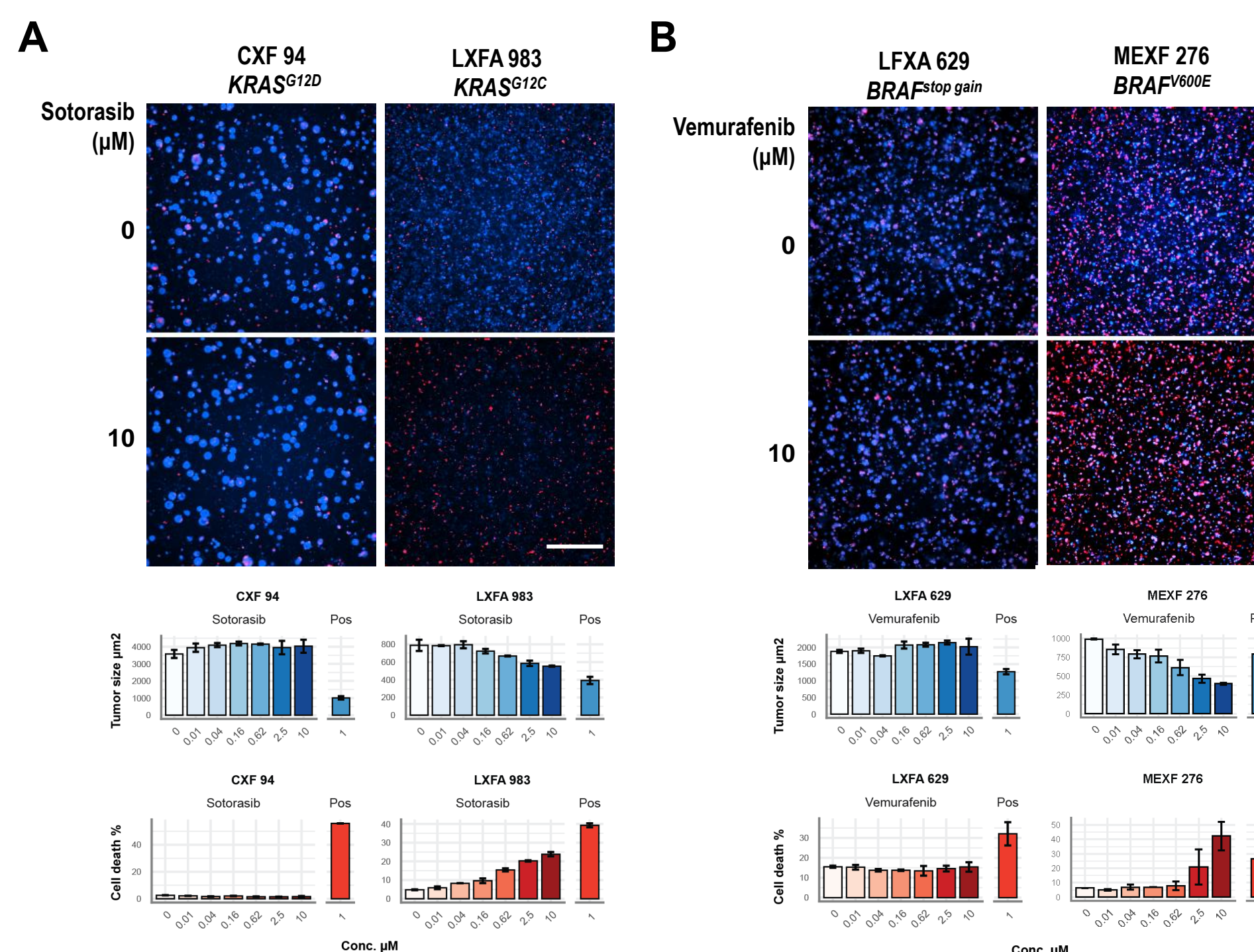


Fig 1. Dose-response profiles of Sotorasib (A) and Vemurafenib (B) with tumor size and the percentage of cell death. The mutation-specific panels were treated with 6 doses of the compound, and Staurosporine 1μM as positive control for 4 days. Subsequently, the 3D models were stained with Hoechst and DRAQ7, and subject to high-content imaging analysis. The results of a non-responder line and responder line from KRAS and BRAF mutation panel are shown. Scale bar = 200μm

Fig 3. High data reproducibility in Cypre 3D platform

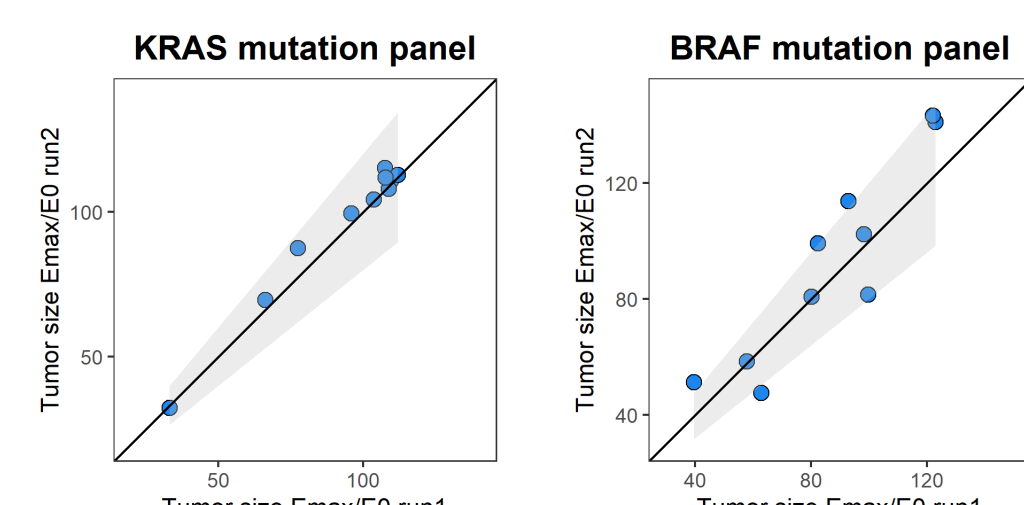


Fig 3. Duplicate testing of BRAF mutation and KRAS mutation panels shows consistent Emax/EO values for tumor size in each 3D tumor model.

Fig 2. The list of each fixed Mutation-specific panel and their response to the targeted therapies

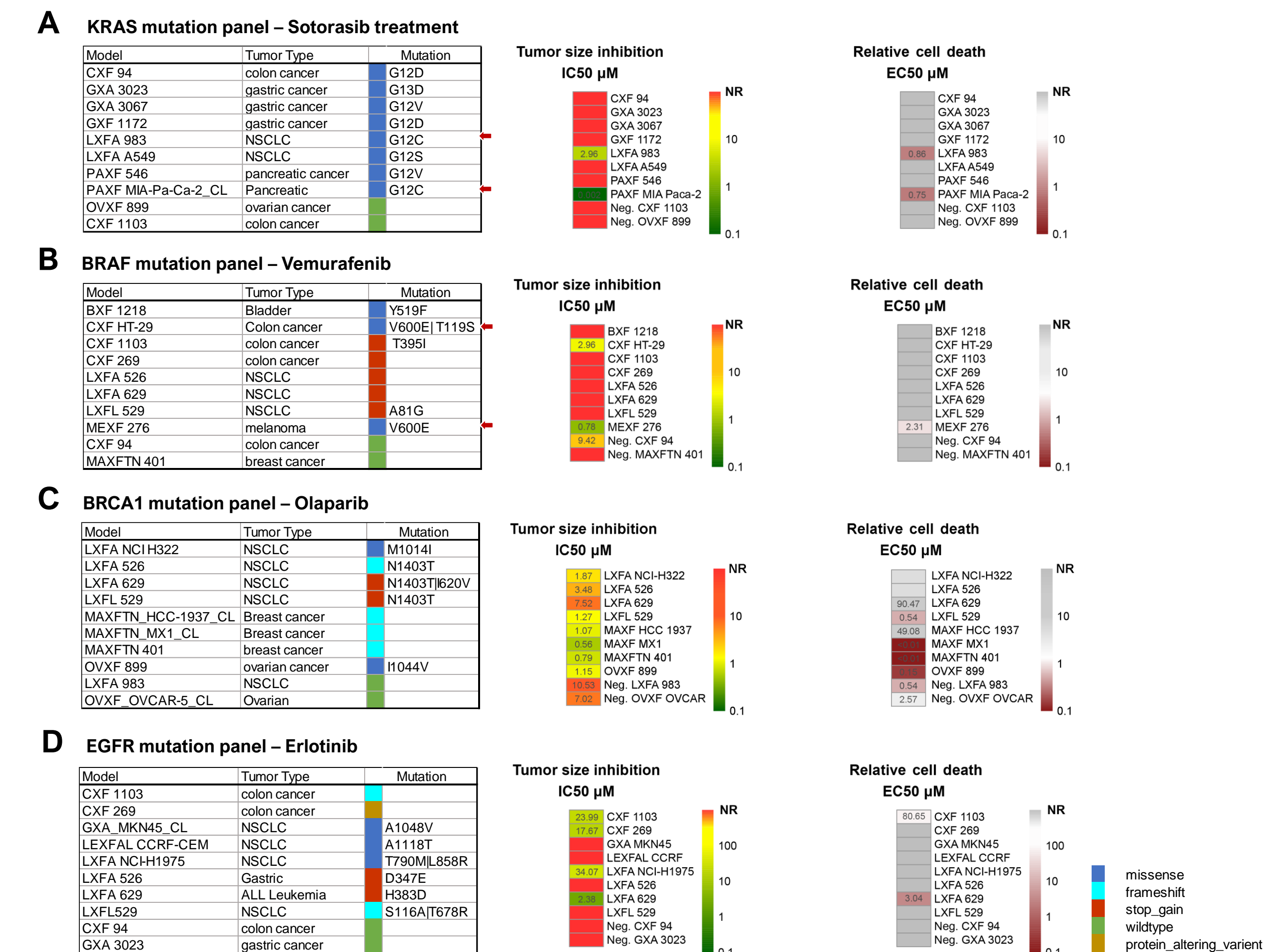


Fig 2. Each Mutation-specific panel is comprised of 8 mutant models and 2 wildtype models across multiple tumor types. KRAS (A), BRAF (B), BRCA1 (C) and EGFR (D) panels were treated with their targeted therapies Sotorasib, Vemurafenib, Olaparib and Erlotinib, respectively. The drug efficacy was visualized using heatmap showing IC50 value of relative tumor size inhibition and EC50 value of cell death induction (comparison to vehicle and positive control).

4 CONCLUSION

- Cypre 3D Mutational Panels comprising PDX/CDX *in vitro* models in 3D hydrogels with fibroblasts screen for anti-tumor effects of targeted compounds.
- Mutational Panels include 10 models expressing either KRAS, BRAF, BRCA1, or EGFR (8 mutant, 2 wildtype).
- Assay endpoints include tumor size reduction and tumor cell death via high content image analysis.
- Responders enabled indication and model selection for subsequent preclinical studies.



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